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Chapter: Emmprin (CD147), a Tumor Cell Surface Inducer of Matrix Metalloproteinase

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## CHAPTER 7

## Emmprin (CD147), a Tumor Cell Surface Inducer of Matrix Metalloproteinase Production

Bryan P. Toole

## Introduction

Emmprin is a member of the Ig superfamily that plays an essential role in several normal tissues but is particularly enriched on the surface of malignant tumor cells in vitro and in vivo. Tumor cell emmprin stimulates production of several matrix metalloproteinases (MMPs) by fibroblasts and endothelial cells, but it also acts in an autocrine fashion to increase MMP synthesis and invasiveness in tumor cells themselves. In addition, emmprin acts as a docking protein for interstitial collagenase on the surface of tumor cells. Increased expression of emmprin in weakly malignant, human breast cancer cells leads to dramatic augmentation of tumor growth and invasion in vivo.

Several important aspects of tumor progression involve proteolytic modification of the pericellular matrix around tumor cells by matrix metalloproteinases (MMPs). For example, MMPs have been implicated in invasion through basement membranes and interstitial matrices, angiogenesis, and tumor cell growth. Strong support for the involvement of MMPs in tumor invasion in vivo comes from experiments in which natural or synthetic inhibitors of MMPs were shown to prevent metastasis in experimental animal models.<sup>1-3</sup> In this chapter I will discuss the function of emmprin, an important regulator of MMP synthesis, in tumor cell invasion.

Emmprin was initially identified as a factor associated with the surface of tumor cells that stimulates synthesis of matrix metalloproteinases by fibroblasts.<sup>4,5</sup> On cloning of emmprin cDNA,<sup>4</sup> it became apparent that emmprin is a member of the Ig superfamily and that it is identical to human basigin<sup>6</sup> and the M6 antigen present on membranes of leukocytes from patients with arthritis,<sup>7</sup> proteins whose functions were not then known. Emmprin is homologous to proteins independently discovered in a wide variety of systems in other species, e.g., mouse gp42 and basigin,<sup>8,9</sup> rat OX47 and CE9,<sup>10,11</sup> and chick 5A11, HT7 and neurothelin.<sup>12-14</sup> Emmprin and its homologs are now also termed CD147. In addition, it is evident that there is a family of molecules related to emmprin. For example, embigin and basigin are closely related,<sup>6</sup> and rat synaptic membranes contain two major Ig superfamily proteins, gp65 and gp55, that are related but not identical to the rat homolog of emmprin.<sup>15</sup>

## Tumor Cell Emmprin Stimulates Fibroblast Production of MMPs

A surprising development with respect to MMP production in tumors was the discovery that most MMPs, e.g., interstitial collagenase (MMP-1), collagenase-3 (MMP-13), gelatinase A (MMP-2), gelatinase B (MMP-9), stromelysin-1 (MMP-3), stromelysin-3 (MMP-11), and

membrane type-MMPs (MT-MMPs), are mainly produced by stromal fibroblasts associated with tumors.<sup>16-20</sup> Moreover, these stromal MMPs contribute significantly to tumor progression in vivo.<sup>20-22</sup> However, MMPs are produced both by stromal cells and by tumor cells, possibly depending on the stage of progression of the tumor, and both sources of MMPs are likely to be important.<sup>17,23,24</sup> Matrilysin (MMP-7) appears to be unique in its restriction to epithelial and carcinoma cells.<sup>17,25</sup>

The almost ubiquitous production of MMPs by stromal cells within tumors, but not within most normal adult tissues, implies that tumor cells may exert regulatory effects on the stromal cells, inducing them to express elevated levels of MMPs. Although it is clear that soluble cytokines and growth factors contribute to this process,<sup>26-28</sup> it is also apparent that tumor cell membrane-bound factors are involved. The first systematic investigation of the latter took place in the laboratory of Dr. Chitra Biswas, where initial experiments suggested that tumor cell-secreted or shed factors were responsible for stimulation of synthesis of MMP-1 by fibroblasts.<sup>29,30</sup> However, subsequent experiments in the Biswas lab showed that most of the MMP-1-stimulatory factor produced by B16 murine melanoma and LX-1 human lung carcinoma cells was plasma membrane-derived, and that this factor could act via direct cell-cell interaction or via shedding of the factor from the cell surface.<sup>31,32</sup> An activity-blocking monoclonal antibody was produced against the factor (originally called tumor cell-derived collagenase stimulatory factor or TCSF)<sup>33</sup> which led to its cloning and full characterization as a transmembrane glycoprotein and member of the Ig superfamily.<sup>4,5</sup> It was also shown to be present in normal tissue<sup>34</sup> and to stimulate production of several MMPs by fibroblasts,<sup>35</sup> and was thus renamed emmprin (extracellular matrix metalloproteinase inducer).<sup>4</sup> (Sadly, Chitra Biswas died in 1993, after having completed the molecular characterization of emmprin).

More recent data has revealed that purified emmprin not only stimulates synthesis of MMPs by fibroblasts but also by endothelial cells. Emmprin stimulates production of interstitial collagenase (MMP-1), gelatinase A (MMP-2) and stromelysin-1 (MMP-3) in both cell types (Refs. 5, 35; Zucker S, Cio J, Rollo EE, Toole BP, unpublished results). Emmprin-mediated stimulation of MMP-1 synthesis in human lung fibroblasts is dependent on the activity of the MAP kinase, p38, but not ERK1/2 or SAPK/JNK.<sup>36</sup> A recent study has shown that emmprin also stimulates synthesis of membrane-type-MMPs (MT-MMPs) in co-cultures of human glioblastoma cells expressing high levels of emmprin with brain tumor-derived fibroblasts.<sup>37</sup> Both MT1- and MT2-MMP were stimulated in this system. Increased activation of MMP-2 by emmprin has also been observed,<sup>35,37</sup> presumably due to the action of MT-MMPs.<sup>38,39</sup> However, it has been noted that different fibroblast populations differ widely in their response to emmprin;<sup>5,35</sup> the basis for this difference has not yet been elucidated.

The effect of emmprin on tumor cell invasion has been examined in co-cultures of oral squamous cell carcinoma cells and peritumor-derived fibroblasts.<sup>40</sup> In this study the tumor cells were plated on a filter coated with reconstituted basement membrane matrix; the fibroblasts were plated in a well beneath the filter. Tumor cell invasion of the matrix was found to be dependent on emmprin and to result from emmprin stimulation of MMP-2 production, presumably by the fibroblasts.<sup>40</sup>

### Autocrine Action of Emmprin Promotes Tumor Cell Invasiveness

Recent data suggest that emmprin acts in an autocrine as well as paracrine fashion. Transfection of weakly malignant MB-MDA436 human breast carcinoma cells with emmprin cDNA leads to an increase in MMP-2 and MT-MMP production (Ref. 41; Caudroy S, Polette M, Nawrocki-Raby B, Toole BP, Zucker S, Birembaut P, submitted for publication). These emmprin-transfected cells were found to be more invasive than vector-transfected controls. Similar findings have been made with the more malignant MDA-435 breast carcinoma cell line without transfection, in that MMP-2 production by and invasiveness of these cells were shown to be emmprin-dependent.<sup>42</sup> In the latter study it was also shown that soluble emmprin inhibits

endogenous emmprin action,<sup>42</sup> most likely between emmprin molecules.<sup>42-44</sup>

### Emmprin Docks MMP-1 on the Cell Surface

After synthesis and secretion, some MMPs, such as MMP-2, bind to either  $\alpha v \beta 3$  integrins or  $\alpha 5 \beta 1$  integrins. Binding of the latter complex leads to activation. Activation involving MT-MMP may occur on the cell surface via interaction with CD44.<sup>45</sup> Docking of MMPs at these docking sites has been seen in recent studies. We have shown that, in addition to docking protein for MMP-1,<sup>49</sup> we show immunocytochemistry that MMP-1 forms a complex with collagenase on the surface of lung carcinoma cells. Since collagenase localization of MMP-1 on the tumor cell

### Emmprin Promotes Tumor Growth

Although it is now apparent that many tumors express the level of emmprin expression in tumors is higher than in corresponding normal tissue.<sup>36,51-53</sup> Emmprin and gelatinase A (MMP-2) are expressed in normal lung tissue vs squamous cell carcinoma vs ductal carcinomas of the breast.<sup>52</sup> Emmprin and the majority of lung carcinomas. Both tumor stromal cells and peritumoral epithelial cells express emmprin. Normal and benign epithelia were not expressed. On the other hand, were restricted to stromal cells close to the tumor. mRNA was also analyzed by Northern blot. Results showed low expression in normal lung tissue. Progression in both lung and breast cancer quantitative image cytometry showed that pre-invasive and invasive nests of tumor cells express emmprin. Both normal and tumor epithelial cells express emmprin. Emmprin was much stronger in tumor tissue than in normal tissue. It is higher in transitional cell carcinomas of the bladder than in malignant glioblastomas than in benign gliomas. It is expressed at a moderately high level in normal oral squamous cell carcinoma is associated with

Since malignant tumor cells often express higher levels than normal and benign cells, we re-examine the role of emmprin in tumor progression.<sup>41</sup> We used growing primary tumors in nude mice and transfected the cells with emmprin cDNA and expression of emmprin. The emmprin transfection controls in monolayer cell culture. The tumor groups of 10 nude mice in three separate experiments. In all three experiments, the mice injected with emmprin cDNA survived for a 12 week period whereas controls grew and died by 8 weeks. In addition, the emmprin transfected mice showed extensive invasion into surrounding abdominal cavity. Survival was markedly decreased with the emmprin transfected mice. We conclude that increased expression of emmprin promotes tumor progression.

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produced by stromal fibroblasts associated with tumor progression, both by stromal cells and by tumor cells, the tumor, and both sources of MMPs are 7) appears to be unique in its restriction to

stromal cells within tumors, but not within normal tissues. It may exert regulatory effects on the stromal cells. Although it is clear that soluble cytokines can also appear that tumor cell membrane-associated MMPs. Investigation of the latter took place in the experiments suggested that tumor cell-secreted factors of synthesis of MMP-1 by fibroblasts.<sup>29,30</sup> b showed that most of the MMP-1-stimulated LX-1 human lung carcinoma cells would act via direct cell-cell interaction or via an activity-blocking monoclonal antibody was a cell-derived collagenase stimulatory factor characterization as a transmembrane glycoprotein known to be present in normal tissue<sup>34</sup> and to its,<sup>35</sup> and was thus renamed emmprin (extracellular matrix metalloproteinase inducer).

Emmprin not only stimulates synthesis of MMPs but also stimulates production of interstitial collagenase (MMP-1) in both cell types (Refs. unpublished results). Emmprin-mediated stimulation is dependent on the activity of the MAP kinase. A recent study has shown that emmprin also stimulates production of MMP-2 by human glioblastoma brain tumor-derived fibroblasts.<sup>37</sup> Both systems. Increased activation of MMP-2 by emmprin due to the action of MT-MMPs.<sup>38,39</sup> How populations differ widely in their response to emmprin has yet been elucidated.

Emmprin has been examined in co-cultures of oral squamous cell carcinoma cells and stromal fibroblasts.<sup>40</sup> In this study the tumor cells invaded the basement membrane matrix; the fibroblasts did not. In the presence of emmprin stimulation of MMP-2 production, pre-

### Emmprin Promotes Tumor Cell Invasiveness

Emmprin acts in both autocrine as well as paracrine fashion. Transfection of breast carcinoma cells with emmprin cDNA increased MMP production (Ref. 41; Caudroy S, Polette M, et al., submitted for publication). These emmprin-transfected cells were more invasive than vector-transfected controls. Similar findings were found in the MDA-435 breast carcinoma cell line without emmprin. Invasiveness of these cells were shown to be dependent on emmprin. It has also been shown that soluble emmprin inhibits

endogenous emmprin action,<sup>42</sup> most likely due to interference with homophilic interactions between emmprin molecules.<sup>42-44</sup>

### Emmprin Docks MMP-1 on the Tumor Cell Surface

After synthesis and secretion, some MMPs bind back to the tumor cell surface. For example, MMP-2 binds to either  $\alpha v \beta 3$  integrin<sup>45</sup> or to a TIMP2-MT-MMP complex; formation of the latter complex leads to activation of MMP-2.<sup>38,39</sup> A similar mechanism of binding and activation involving MT-MMP may occur with collagenase-3.<sup>46</sup> Gelatinase B can bind to the cell surface via interaction with CD44<sup>47</sup> or a component of collagen type IV.<sup>48</sup> Presentation of MMPs at these docking sites has been shown to promote tumor cell invasiveness.<sup>38,45,47</sup> In a recent study we have shown that, in addition to stimulating MMP production, emmprin is a docking protein for MMP-1.<sup>49</sup> We showed by phage display, affinity chromatography and immunocytochemistry that MMP-1 forms a complex with emmprin on the surface of human lung carcinoma cells. Since collagenase activity is essential for invasion of fibrous tissues,<sup>50</sup> localization of MMP-1 on the tumor cell surface would facilitate this process.

### Emmprin Promotes Tumor Growth and Invasion In Vivo

Although it is now apparent that many normal embryonic and adult tissues express emmprin, the level of emmprin expression in tumors, especially malignant tumors, is usually much greater than in corresponding normal tissue.<sup>36,51-55</sup> For example, in one study, the relative distribution of emmprin and gelatinase A (MMP-2) mRNAs was compared by *in situ* hybridization in normal lung tissue vs squamous cell carcinomas of the lung and in benign mammary growths vs ductal carcinomas of the breast.<sup>52</sup> Emmprin mRNA was detected in all breast carcinomas and the majority of lung carcinomas. Both pre-invasive and invasive cancer cells were positive, but tumor stromal cells and peritumoral tissue showed insignificant emmprin mRNA reactivity. Normal and benign epithelia were negative. MMP-2 and MMP-1 mRNAs, on the other hand, were restricted to stromal cells close to tumor clusters.<sup>36,52</sup> The expression of emmprin mRNA was also analyzed by Northern blots which were then densitometrically scanned; the results showed low expression in normal or benign tissues but high levels at all stages of tumor progression in both lung and breast cancers.<sup>52</sup> Analyses of distribution within tumors made by quantitative image cytometry showed that high levels of emmprin mRNA were expressed in pre-invasive and invasive nests of tumor cells versus low amounts in normal or peritumoral tissues.<sup>52</sup> Both normal and tumor epithelia stained with antibody to emmprin, but expression of emmprin was much stronger in tumor tissue.<sup>53</sup> In other studies, emmprin levels were shown to be higher in transitional cell carcinomas of the bladder than in normal bladder epithelium,<sup>51</sup> and in malignant glioblastomas than in benign gliomas and normal brain tissue.<sup>55</sup> Although emmprin is expressed at a moderately high level in normal non-neoplastic keratinocytes,<sup>34</sup> its presence in oral squamous cell carcinoma is associated with MMP production and tumor cell invasion.<sup>40</sup>

Since malignant tumor cells often express emmprin *in vivo* and *in vitro* at much higher levels than normal and benign cells, we recently tested whether over-expression of emmprin stimulates tumor progression.<sup>41</sup> We used human breast carcinoma cells that produce slow-growing primary tumors in nude mice and express relatively low levels of emmprin. We transfected the cells with emmprin cDNA and selected stable transfectant clones with increased expression of emmprin. The emmprin transfectants grew at similar rates to vector-transfected controls in monolayer cell culture. The tumor cells were injected into the mammary fat pad of groups of 10 nude mice in three separate *in vivo* experiments using different transfectant clones. In all three experiments, the mice injected with emmprin transfectants grew large tumors over a 12 week period whereas controls grew small tumors that were primarily detectable only at autopsy. In addition, the emmprin transfectants gave rise to high levels of MMP expression and to extensive invasion into surrounding abdominal wall muscle whereas controls did not. Mouse survival was markedly decreased with the emmprin transfectants compared to controls.<sup>41</sup> We conclude that increased expression of emmprin leads to increased malignant tumor behavior.

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## The Functions of Emmprin are Diverse

Recently, a knockout mouse has been produced in which basigin, the murine homolog of emmprin, is lacking.<sup>56</sup> The null mutant is in most cases unable to undergo implantation, possibly due to the involvement of MMPs in this process.<sup>57,58</sup> However embryos that successfully implant and survive past birth have deficiencies in spermatogenesis,<sup>56,59</sup> retinal and photoreceptor development and maintenance,<sup>60,61</sup> other sensory functions,<sup>62</sup> and lymphocyte responses.<sup>62</sup> Any relevance of MMP stimulation to these latter processes has not been established.

Structural analyses have demonstrated that the transmembrane and cytoplasmic domains of emmprin are highly conserved among species, suggesting that these regions are of functional importance. The properties of the transmembrane region also suggest that intramembrane interactions with other proteins are likely to occur.<sup>6,7,10</sup> Emmprin interacts with integrins,  $\alpha 3 \beta 1$  and  $\alpha 6 \beta 1$ , within the plasma membrane of HT1080 fibrosarcoma cells.<sup>63</sup> It acts as a chaperone for assembly of lactate transporters in the plasma membrane.<sup>64</sup> It binds to cyclophilin A, facilitating HIV virus entry into cells.<sup>65</sup> These interactions are likely to involve the transmembrane and/or cytoplasmic domains of emmprin. Again, however, it is not known whether proteolytic processes stimulated by emmprin are involved in any of these processes. Rather, it seems likely that emmprin has multiple functions, but the underlying mechanisms are presently unknown.

## Conclusions

Increasingly, evidence is appearing that firmly establishes the importance of the stroma in carcinoma progression.<sup>66-69</sup> We propose that interactions of tumor cells and stromal cells lead to synthesis and activation of MMPs that in turn promote tumor invasiveness and that emmprin is a crucial component of these interactions. However, emmprin on the tumor cell surface also appears to be directly involved in tumor cell invasiveness, without stromal interactions, by autocrine stimulation of MMP synthesis and by docking of MMP-1 to the cell surface. It is becoming increasingly apparent that tumor cells create a pericellular environment in which many MMPs and other proteases become concentrated, thereby enhancing the ability of tumor cells to invade extracellular matrices and to process locally precursors of factors that promote tumor progression. Emmprin stimulation of MMP production could play a central role in these processes. However, emmprin is also involved in other pathological and physiological events that may or may not involve regulation of MMP synthesis. Whether or not emmprin serves more than one molecular function in malignant tumor cell behavior remains to be seen.

## References

- Khokha R. Suppression of the tumorigenic and metastatic abilities of murine B16-F10 melanoma cells in vivo by the overexpression of the tissue inhibitor of the metalloproteinases-1. *J Natl Cancer Inst* 1994; 86: 299-304.
- Sledge GW, Qulali M, Goulet R, Bone EA, Fife R. Effect of matrix metalloproteinase inhibitor batimastat on breast cancer regrowth and metastasis in athymic mice. *J Natl Cancer Inst* 1995; 87:1546-1550.
- Hua J, Muschel R. Inhibition of matrix metalloproteinase 9 expression by a ribozyme blocks metastasis in a rat fibrosarcoma model system. *Cancer Res* 1996; 56:5279-5284.
- Biswas C, Zhang Y, DeCastro R, Guo H, Nakamura T et al. The human tumor cell-derived collagenase stimulatory factor (renamed EMMPRIN) is a member of the immunoglobulin superfamily. *Cancer Res* 1995; 55:434-439.
- Guo H, Zucker S, Gordon MK, Toole BP, Biswas C. Stimulation of matrix metalloproteinase production by recombinant extracellular matrix metalloproteinase inducer from transfected chinese hamster ovary cells. *J Biol Chem* 1997; 272:24-27.
- Miyauchi T, Masuzawa Y, Muramatsu T. The basigin group of the immunoglobulin superfamily: Complete conservation of a segment in and around transmembrane domains of human and mouse basigin and chick HT7 antigen. *J Biochem* 1991; 110:770-774.
- Kasinerk W, Fiebigler E, Steffanova J, Baumruker T, Knapp W et al. Human leukocyte activation antigen M6, a member of the Ig superfamily, is the species homologue of rat OX-47, mouse basigin, and chicken HT7 molecule. *J Immunol* 1992; 149:847-854.
- Altruda F, Cervella P, Gaeta ML. Membrane glycoprotein (gp42): Sh and neural-cell adhesion molecules.
- Miyauchi T, Kanekura T, Yamaoka. tributed member of the immunoglobulin V domain and the  $\beta$ -ch J Biochem 1990; 107:316-323.
- Fossum S, Mallert S, Barclay AN. 7 superfamily with an unusual transmembrane domain.
- Nehme CL, Cesario MM, Myies D. compartmentalizes transmembrane main of the rat spermatozoon. *J Cell Biol* 1991; 113:111-121.
- Fadool JM, Linser PJ. 5A11 antigenic interactions in avian neural retina.
- Schlosshauer B, Bauch H, Frank R. actin colocalization. *Europ J Cell Biol* 1991; 111:111-121.
- Seulberger H, Unger CM, Risau W. for one developmentally regulated in helium, epithelial tissue barriers and
- Langnaese K, Beesley PW, Gundelfin. new members of the immunoglobulin superfamily.
- DeClerck YA. Interactions between the extracellular matrix by metalloproteinases.
- Heppner KJ, Matrisian LM, Jensen J. family members in breast cancer regression. *Cancer Res* 1991; 51:273-282.
- Johnsen M, Lund LR, Romer J, A. Common themes in proteolytic matrix metalloproteinases.
- Okada A, Bellocq JP, Rouyer N. metalloproteinase (MT-MMP) gene and neck carcinomas. *Proc Natl Acad Sci USA* 1995; 92:1535-1541.
- Masson R, Lefebvre O, Noel A, E. stromelysin-3 metalloproteinase controls tumor growth. *Cell Biol* 1998; 140:1535-1541.
- Itoh T, Tanioka M, Yoshida H, Yoshida T. progression in gelatinase A-deficient mice.
- Sternlicht MD, Lochter A, Sympon. stromelysin-1 promotes mammary carcinoma growth.
- Wright JH, McDonnell S, Portella. tumor cell expression of stromelysin-1 in carcinomas during mouse skin tumor progression.
- Sehgal G, Hua J, Bernhard EJ. metalloproteinase-9 (gelatinase B) expression in human breast carcinomas. *Am J Pathol* 1998; 152:591-596.
- Wilson CL, Heppner KJ, Rudolph I. entially expressed by epithelial cells in human breast carcinomas.
- Ito A, Nakajima S, Sasaguri Y, Nag. MCF-7 cells and human dermal fibroblasts. *Int J Cancer* 1997; 71:112-117.
- Uria JA, Stahle-Backdal M, Seiki M. sion in human breast carcinomas is regulated by tumor cells.
- Westermarck J, Li S, Jaakkola P, Kallunki T. nase-1 expression by tumor cells of human breast carcinomas.
- Biswas C. Tumor cell stimulation of collagenase production. *Commun Cell Tissue Res* 1982; 109:1026-1034.
- Biswas C. Collagenase stimulation in human breast carcinomas. *Cancer Letters* 1984; 24:201-207.

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ric abilities of murine B16-F10 melanoma  
of the metalloproteinases-1. *J Natl Cancer*

ffect of matrix metalloproteinase inhibitor  
athymic mice. *J Natl Cancer Inst* 1995;

ase 9 expression by a ribozyme blocks me-  
1996; 56:5279-5284.

T et al. The human tumor cell-derived  
a member of the immunoglobulin super-

Stimulation of matrix metalloproteinase  
proteinase inducer from transfected chinese

group of the immunoglobulin superfamily:  
ismembrane domains of human and mouse  
770-774.

napp W et al. Human leukocyte activation  
es homologue of rat OX-47, mouse basigin,  
-854.

8. Altruda F, Cervella P, Gaeta ML, Daniele A, Giancotti F et al. Cloning of cDNA for a mouse membrane glycoprotein (gp42): Shared identity to histocompatibility antigens, immunoglobulins and neural-cell adhesion molecules. *Gene* 1989; 85:445-452.
9. Miyauchi T, Kanekura T, Yamaoka A, Ozawa M, Miyazawa S et al. Basigin, a new, broadly distributed member of the immunoglobulin superfamily, has strong homology with both the immunoglobulin V domain and the  $\beta$ -chain of major histocompatibility complex class II anti gen. *J Biochem* 1990; 107:316-323.
10. Fossum S, Mallett S, Barclay AN. The MRC OX-47 antigen is a member of the immunoglobulin superfamily with an unusual transmembrane sequence. *Eur J Immunol* 1991; 21:671-679.
11. Nehme CL, Cesario MM, Myles DG, Koppel DE, Bartles JR. Breaching the diffusion barrier that compartmentalizes transmembrane glycoprotein CE9 to the posterior-tail plasma membrane domain of the rat spermatozoon. *J Cell Biol* 1993; 120:687-694.
12. Fadool JM, Linser PJ. SA11 antigen is a cell recognition molecule which is involved in neuronal-glial interactions in avian neural retina. *Dev Dynamics* 1993; 196:252-262.
13. Schlosshauer B, Bauch H, Frank R. Neurothelin: Amino acid sequence, cell surface dynamics and actin colocalization. *Europ J Cell Biol* 1995; 68:159-166.
14. Seilberger H, Unger CM, Risau W. HT7, neurothelin, basigin, gp42 and OX-47—Many names for one developmentally regulated immunoglobulin-like surface glycoprotein on blood-brain endothelium, epithelial tissue barriers and neurons. *Neurosci Lett* 1992; 140:93-97.
15. Langnaese K, Beesley PW, Gundelfinger ED. Synaptic membrane glycoproteins gp65 and gp55 are new members of the immunoglobulin superfamily. *J Biol Chem* 1997; 272:821-827.
16. DeClerck YA. Interactions between tumour cells and stromal cells and proteolytic modification of the extracellular matrix by metalloproteinases in cancer. *Europ J Cancer* 2000; 36:1258-1268.
17. Fleppner KJ, Matrisian LM, Jensen RA, Rodgers WH. Expression of most matrix metalloproteinase family members in breast cancer represents a tumor-induced host response. *Amer J Pathol* 1996; 149:273-282.
18. Johnsen M, Lund LR, Romer J, Almholt K, Dano K. Cancer invasion and tissue remodeling: Common themes in proteolytic matrix degradation. *Current Opin Cell Biol* 1998; 10:667-671.
19. Okada A, Bellocq JP, Rouyer N, Chenard MP, Rio MC et al. Membrane-type matrix metalloproteinase (MT-MMP) gene is expressed in stromal cells of human colon, breast, and head and neck carcinomas. *Proc Nat Acad Sci USA* 1995; 92:2730-2734.
20. Masson R, Lefebvre O, Noel A, El Fahime M, Chenard MP et al. In vivo evidence that the stromelysin-3 metalloproteinase contributes in a paracrine manner to epithelial cell malignancy. *J Cell Biol* 1998; 140:1535-1541.
21. Itoh T, Tanioka M, Yoshida H, Yoshioka T, Nishimoto H et al. Reduced angiogenesis and tumor progression in gelatinase A-deficient mice. *Cancer Res* 1998; 58:1048-1051.
22. Sternlicht MD, Lochter A, Sympon CJ, Huey B, Rouzier JP et al. The stromal proteinase MMP3/stromelysin-1 promotes mammary carcinogenesis. *Cell* 1999; 98:137-146.
23. Wright JH, McDonnell S, Portella G, Bowden GT, Balmain A et al. A switch from stromal to tumor cell expression of stromelysin-1 mRNA associated with the conversion of squamous to spindle carcinomas during mouse skin tumor progression. *Mol Carcinogenesis* 1994; 10:207-215.
24. Sehgal G, Hua J, Bernhard EJ, Sehgal I, Thompson TC et al. Requirement for matrix metalloproteinase-9 (gelatinase B) expression in metastasis by murine prostatic carcinoma. *Amer J Pathol* 1998; 152:591-596.
25. Wilson CL, Heppner KJ, Rudolph LA, Matrisian LM. The metalloproteinase matrilysin is preferentially expressed by epithelial cells in a tissue-restricted pattern in the mouse. *Mol Biol Cell* 1995; 6:851-869.
26. Ito A, Nakajima S, Sasaguri Y, Nagase H, Mori Y. Co-culture of human breast adenocarcinoma MCF-7 cells and human dermal fibroblasts enhances the production of matrix metalloproteinases 1,2 and 3 in fibroblasts. *Brit J Cancer* 1995; 71:1039-1045.
27. Uria JA, Stahle-Backdal M, Seiki M, Fuyo A, Lopez-Otin C. Regulation of collagenase-3 expression in human breast carcinomas is mediated by stromal-epithelial cell interactions. *Cancer Res* 1997; 57:4882-4888.
28. Westermarck J, Li S, Jaakkola P, Kallunki T, Grenman R et al. Activation of fibroblast collagenase-1 expression by tumor cells of squamous cell carcinomas is mediated by p38 mitogen-activated protein kinase and c-jun NH<sub>2</sub>-terminal kinase-2. *Cancer Res* 2000; 60:7156-7162.
29. Biswas C. Tumor cell stimulation of collagenase production by fibroblasts. *Biochem Biophys Res Commun* 1982; 109:1026-1034.
30. Biswas C. Collagenase stimulation in cocultures of human fibroblasts and human tumor cells. *Cancer Letters* 1984; 24:201-207.

BEST AVAILABLE COPY

31. Biswas C, Nugent MA. Membrane association of collagenase stimulatory factor(s) from B-16 melanoma cells. *J Cell Biochem* 1987; 35:247-258.
32. Nabeshima K, Lane WS, Biswas C. Partial sequencing and characterization of the tumor cell-derived collagenase stimulatory factor. *Arch Biochem Biophys* 1991; 285:90-96.
33. Ellis SM, Nabeshima KN, Biswas C. Monoclonal antibody preparation and purification of a tumor cell collagenase-stimulatory factor. *Cancer Res* 1989; 49:3385-3391.
34. DeCastro R, Zhang Y, Guo H, Kataoka H, Gordon MK et al. Human keratinocytes express EMMPRIN, an extracellular matrix metalloproteinase inducer. *J Invest Dermatol* 1996; 106:1260-1265.
35. Kataoka H, DeCastro R, Zucker S, Biswas C. The tumor cell-derived collagenase stimulatory factor, TCSF, increases expression of interstitial collagenase, stromelysin and 72-kDa gelatinase. *Cancer Res* 1993; 53:3154-3158.
36. Lim M, Martinez T, Jablons D, Cameron R, Guo H et al. Tumor-derived EMMPRIN (extracellular matrix metalloproteinase inducer) stimulates collagenase transcription through MAPK p38. *FEBS Lett* 1998; 441:88-92.
37. Sameshima T, Nabeshima K, Toole BP, Yokogami K, Okada Y et al. Glioma cell extracellular matrix metalloproteinase inducer (EMMPRIN) (CD147) stimulates production of membrane-type matrix metalloproteinases and activated gelatinase A in co-cultures with brain-derived fibroblasts. *Cancer Lett* 2000; 157:177-184.
38. Nakahara H, Howard L, Thompson EW, Sato H, Seiki M et al. Transmembrane/cytoplasmic domain-mediated membrane type 1-matrix metalloproteinase docking to invadopodia is required for invasion. *Proc Natl Acad Sci USA* 1997; 94:7959-7964.
39. Zucker S, Drews M, Conner C, Foda HD, DeClerck YA et al. Tissue inhibitor of metalloproteinase-2 (TIMP-2) binds to the catalytic domain of the cell surface receptor, membrane type 1-matrix metalloproteinase 1 (MT1-MMP). *J Biol Chem* 1998; 273:1216-1222.
40. Bordador LC, Li X, Toole BP, Chen B, Regezi J et al. Expression of emmprin by oral squamous cell carcinoma. *Int J Cancer* 2000; 85:347-352.
41. Zucker S, Hymowitz M, Rollo EE, Mann R, Conner CE et al. Tumorigenic potential of extracellular matrix metalloproteinase inducer (EMMPRIN). *Amer J Pathol*; in press.
42. Sun J, Hemler ME. Regulation of MMP-1 and MMP-2 production through CD147/extracellular matrix metalloproteinase inducer interactions. *Cancer Res* 2001; 61:2276-2281.
43. Fadoo JM, Linser PJ. Evidence for the formation of multimeric forms of the 5A11/HT7 antigen. *Biochem Biophys Res Commun* 1996; 229:280-286.
44. Yoshida S, Shibata M, Yamamoto S, Hagihara M, Asai N et al. Homo-oligomer formation by basigin, an immunoglobulin superfamily member, via its N-terminal immunoglobulin domain. *Europ J Biochem* 2000; 267:4372-4380.
45. Brooks PC, Stromblad S, Sanders LC, von Schalscha TL, Aimes RT et al. Localization of matrix metalloproteinase MMP-2 to the surface of invasive cells by interaction with integrin alpha V beta 3. *Cell* 1996; 85:683-693.
46. Knauper V, Will H, Lopez-Otin C, Smith B, Atkinson SJ et al. Cellular mechanisms for human procollagenase-3 (MMP-13) activation. Evidence that MT1-MMP (MMP-14) and gelatinase A (MMP-2) are able to generate active enzyme. *J Biol Chem* 1996; 271:17124-17131.
47. Yu Q, Stamenkovic I. Cell surface-localized matrix metalloproteinase-9 proteolytically activates TGF-beta and promotes tumor invasion and angiogenesis. *Genes Dev* 2000; 14:163-176.
48. Olson MW, Torch M, Gervasi DC, Sado Y, Ninomiya Y et al. High affinity binding of latent matrix metalloproteinase-9 to the alpha2(IV) chain of collagen IV. *J Biol Chem* 1998; 273:10672-10681.
49. Guo H, Li R, Zucker S, Toole BP. EMMPRIN (CD147), an inducer of matrix metalloproteinase synthesis, also binds interstitial collagenase to the tumor cell surface. *Cancer Res* 2000; 60:888-891.
50. Benbow U, Schoenemark MP, Mitchell TI, Rutter JL, Shimokawa K et al. A novel host/tumor cell interaction activates matrix metalloproteinase 1 and mediates invasion through type 1 collagen. *J Biol Chem* 1999; 274:25371-25378.
51. Muraoka K, Nabeshima K, Murayama T, Biswas C, Kono M. Enhanced expression of a tumor cell-derived collagenase-stimulatory factor in urothelial carcinoma: Its usefulness as a tumor marker for bladder cancers. *Int J Cancer* 1993; 55:19-26.
52. Polette M, Gilles C, Marchand V, Lorenzato M, Toole B et al. Tumor collagenase stimulatory factor (TCSF) expression and localization in human lung and breast cancers. *J Histochem Cytochem* 1997; 45:703-710.
53. Caudroy S, Polette M, Tournier JM, Burlet H, Toole B et al. Expression of the extracellular matrix metalloproteinase inducer (EMMPRIN) and the matrix metalloproteinase-2 in bronchopulmonary and breast lesions. *J Histochem Cytochem* 1999; 47:1575-1580.
54. Dalberg K, Eriksson E, Enberg U. Matrix metalloproteinase, and extracellular matrix. Correlation with invasive growth of human melanoma cells. *Int J Cancer* 2000; 88:21-27.
55. Sameshima T, Nabeshima K, Toole B (CD147), a cell surface inducer of matrix metalloproteinase. *Int J Cancer* 2000; 88:21-27.
56. Igakura T, Kadomatsu K, Kaname T, an immunoglobulin superfamily member and spermatogenesis. *Dev Biol* 1997; 191:1-10.
57. Alexander CM, Hansell EJ, Behrendt function of matrix metalloproteinases during mouse embryo implantation. *Dev Biol* 2000; 221:213-2133.
58. Vu TH, Werb Z. Matrix metalloproteinases and the regulation of tissue remodeling. *Nature* 2000; 403:910-916.
59. Toyama Y, Maekawa M, Kadomatsu K. Role of defective spermatogenesis in the development of the testis. *Dev Biol* 1997; 191:1-10.
60. Hori K, Katayama N, Kachi S. Cone deficiency. *Invest Ophthalmol Vis Sci* 1997; 38:2133-2133.
61. Ochritsch JD, Moroz TM, Kadomatsu failed photoreceptor maturation in 5A11/HT7 antigen. *Int J Cancer* 2000; 85:347-352.
62. Igakura T, Kadomatsu K, Taguchi O, of the immunoglobulin superfamily, in blood-brain barrier. *Biochem Biophys Res Commun* 1996; 229:280-286.
63. Berdichevski F, Chang S, Bodorova J. Associated proteins: Evidence that a3b1 is associated with the cell surface receptor. *Int J Cancer* 2000; 85:347-352.
64. Kirk P, Wilson MC, Heddl C, Brown lactate transporters MCT1 and MCT4. *Am J Physiol* 1998; 275:R1039-R1044.
65. Pushkarsky T, Zybath G, Dubrovsky I. Infection by interacting with virus. *Int J Cancer* 2000; 85:347-352.
66. Olumi AF, Grossfeld GD, Hayward fibroblasts direct tumor progression of prostate cancer. *Proc Natl Acad Sci USA* 1999; 96:5002-5007.
67. Park CC, Bissell MJ, Barcellos-Hoff malignant phenotype. *Mol Med Today* 1999; 24:52-57.
68. Moirand F, Man YG, Arnould L, Bratt genetic alterations in the stromal and epithelial cells. *Cancer Res* 2000; 60:2562-2567.
69. Shekhar MP, Werdell J, Santner SJ, Pa role in breast epithelial growth and progression. *Cancer Res* 2001; 61:1320-1325.

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- genase stimulatory factor(s) from B-16 melanoma cells. *Cell Invasion and Characterization of the Tumor Cell Body Preparation and Purification of a Tumor Cell-Derived Collagenase Stimulatory Factor*. *Int J Cancer* 1991; 285:90-96.
- on MK et al. Human keratinocytes express a cell surface inducer of matrix metalloproteinases. *J Invest Dermatol* 1996; 106:1260-1265.
- umor cell-derived collagenase stimulatory factor, stromelysin and 72-kDa gelatinase. *Cancer Res* 1996; 56:1260-1265.
- et al. Tumor-derived EMPRIN (extracellular matrix metalloproteinase inducer) transcription through MAPK p38. *FEBS Lett* 1997; 401:126-130.
- K, Okada Y et al. Glioma cell extracellular matrix metalloproteinase inducer (CD147) stimulates production of membrane-type 1 matrix metalloproteinase in co-cultures with brain-derived fibroblasts. *Int J Cancer* 1997; 72:1216-1222.
- iki M et al. Transmembrane/cytoplasmic domain docking to invadopodia is required for cell invasion. *Proc Natl Acad Sci USA* 1999; 96:1216-1221.
- YA et al. Tissue inhibitor of metalloproteinase-1 surface receptor, membrane type 1-matrix metalloproteinase-1. *J Biol Chem* 1996; 271:1216-1222.
- al. Expression of emmprin by oral squamous cell carcinoma. *Am J Pathol* in press.
- IP-2 production through CD147/extracellular matrix metalloproteinase inducer (CD147) multimeric forms of the 5A11/HT7 antigen. *Int J Cancer* 2001; 61:2276-2281.
- Asai N et al. Homo-oligomer formation by the N-terminal immunoglobulin domain. *Europ J Cell Biol* 1999; 85:121-126.
- a TL, Aimes RT et al. Localization of matrix metalloproteinase-9 proteolytically activates TGF- $\beta$ 1. *Genes Dev* 2000; 14:163-176.
- niya Y et al. High affinity binding of latent TGF- $\beta$ 1 to CD147. *J Biol Chem* 1998; 273:17124-17131.
- CD147, an inducer of matrix metalloproteinase-9. *Cancer Res* 2000; 60:888-891.
- JL, Shimokawa K et al. A novel host/tumor cell surface inducer of matrix metalloproteinase-1 mediates invasion through type 1 collagen. *Cancer Res* 1999; 59:5002-5011.
- , Koono M. Enhanced expression of a tumor cell surface inducer of matrix metalloproteinase-9 in human carcinoma: Its usefulness as a tumor marker. *Cancer Res* 2000; 60:324-329.
- Toole B et al. Tumor collagenase stimulatory factor and breast cancers. *J Histochem Cytochem* 1999; 47:1575-1580.
- Toole B et al. Expression of the extracellular matrix metalloproteinase-2 in bronchopulmonary adenocarcinoma. *Int J Cancer* 1999; 82:1216-1222.
54. Dalberg K, Eriksson E, Enberg U, Kjellman M, Backdahl M. Gelatinase A, membrane type 1 matrix metalloproteinase, and extracellular matrix metalloproteinase inducer mRNA expression: Correlation with invasive growth of breast cancer. *World J Surgery* 2000; 24:334-340.
55. Sameshima T, Nabeshima K, Toole BP, Yokogami K, Okada Y et al. Expression of EMPRIN (CD147), a cell surface inducer of matrix metalloproteinases, in normal human brain and gliomas. *Int J Cancer* 2000; 88:21-27.
56. Igakura T, Kadomatsu K, Kaname T, Muramatsu H, Fan QW et al. A null mutation in basigin, an immunoglobulin superfamily member, indicates its important roles in peri-implantation development and spermatogenesis. *Dev Biol* 1998; 194:152-165.
57. Alexander CM, Hansell EJ, Behrendtsen C, Flannery ML, Kishnani NS et al. Expression and function of matrix metalloproteinases and their inhibitors at the maternal-embryonic boundary during mouse embryo implantation. *Development* 1996; 122:1723-1736.
58. Vu TH, Werb Z. Matrix metalloproteinases: effectors of development and normal physiology. *Genes Dev* 2000; 14:2123-2133.
59. Toyama Y, Mackawa M, Kadomatsu K, Miyauchi T, Muramatsu T et al. Histological characterization of defective spermatogenesis in mice lacking the basigin gene. *Anat Histol Embryol* 1999; 128:205-213.
60. Hori K, Katayama N, Kachi S, Kondo M, Kadomatsu K et al. Retinal dysfunction in basigin deficiency. *Invest Ophthalmol Vis Sci* 2000; 41:3128-3133.
61. Ochrietor JD, Moroz TM, Kadomatsu K, Muramatsu T, Linser PJ. Retinal degeneration following failed photoreceptor maturation in 5A11/basigin null mice. *Exp Eye Res* 2001; 72:467-477.
62. Igakura T, Kadomatsu K, Taguchi O, Muramatsu H, Kaname T et al. Roles of basigin, a member of the immunoglobulin superfamily, in behavior as to an irritating odor, lymphocyte response and blood-brain barrier. *Biochem Biophys Res Commun* 1996; 224:33-36.
63. Berditschewski F, Chang S, Bodorova J, Hemler ME. Generation of monoclonal antibodies to integrin-associated proteins: Evidence that  $\alpha 3 \beta 1$  complexes with EMPRIN/basigin/OX47/M6. *J Biol Chem* 1997; 272:29174-29180.
64. Kirk P, Wilson MC, Heddle C, Brown MH, Barclay AN et al. CD147 is tightly associated with lactate transporters MCT1 and MCT4 and facilitates their cell surface expression. *EMBO J* 2000; 19:3896-3904.
65. Pushkarsky T, Zybath G, Dubrovsky L, Yurchenko V, Guo H et al. CD147 facilitates HIV-1 infection by interacting with virus-associated cyclophilin A. *Proc Natl Acad Sci USA* 2001; 98:6360-6365.
66. Olumi AF, Grossfeld GD, Hayward SW, Carroll PR, Tlsty TD et al. Carcinoma-associated fibroblasts direct tumor progression of initiated human prostatic epithelium. *Cancer Res* 1999; 59:5002-5011.
67. Park CC, Bissell MJ, Barcellos-Hoff MH. The influence of the microenvironment on the malignant phenotype. *Mol Med Today* 2000; 6:324-329.
68. Moiraf F, Man YG, Arnould L, Bratthauer GL, Ratschek M et al. Concurrent and independent genetic alterations in the stromal and epithelial cells of mammary carcinoma: Implications for tumorigenesis. *Cancer Res* 2000; 60:2562-2566.
69. Shekhar MP, Werdell J, Santner SJ, Pauley RJ, Tait L. Breast stroma plays a dominant regulatory role in breast epithelial growth and differentiation: Implications for tumor development and progression. *Cancer Res* 2001; 61:1320-1326.

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